

# Potential Probiotic Analysis: Indigenous Lactic Acid Bacteria from Freshly Drawn Goat Milk

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**Abstract**— The raw goat milk is a good source of potential probiotic microorganisms. In developing countries, the goat milk was consumed widely because of its therapeutic and nutritional values. The potential probiotic bacteria has some properties such as, survival in the gastrointestinal conditions like low pH and bile tolerance, production of antimicrobial compounds and adhesion to intestinal mucosa. Thirty Lactic acid bacteria (LAB) isolated from goat milk in Tirupur and Erode region. Among 30 isolates, the nine isolates were selected for studying the characterization of potential probiotic of LAB. So the screening process were carried out like cell hydrophobicity, phenol resistance, antioxidant assay, cholesterol reducing activity, auto and co-aggregation, acid and bile tolerance. The results obtained showed that the isolates G13 has high spectrum of action. This isolate were undergo to detect the bacteriocin activity, column chromatography and SDS-PAGE. G13 isolates showed inhibitory activity in well-diffusion assay when tested against *E.coli* and *S.aureus*. So, those inhibitory substances are characterized as antimicrobial proteins called bacteriocins. Further identification of bacteriocin produced by G13 was confirmed by SDS-PAGE, where it shows the low molecular weight protein (17 kDa).

**Keywords**— potential probiotic, cell hydrophobicity, phenol resistance, antioxidant assay, SDS-PAGE

## I. INTRODUCTION

Probiotics are defined as “living micro-organisms, which upon ingestion in certain numbers exert health benefits beyond inherent basic nutrition”. FAO/WHO has adopted the definition of probiotics as “live microorganisms which when administered in adequate amounts confer a health benefit on the host” [1]. To achieve a probiotic status, microorganism must fulfil a number of criteria related to safety, functional effects and technological properties [2].

Lactic acid bacteria were utilized as natural preservatives in fermentation industry. It has advantageous health issue on host but the most important method is selection of strain based on physiological properties [3]. Lactic acid bacteria have the capacity to prevent the gastrointestinal tract from the pathogens. For pancreatic enzymes and other gastric secretions genus *Lactobacillus* show more resistant [4].

Ingestion of fermented goat milk amended anti-atherogenicity in healthy cases: it sustained resistance of the lipoprotein fraction to oxidation, lowered levels of per oxidized lipoproteins, oxidized LDL, 8-isoprostanes and glutathione redox ratio and enhanced total antioxidant activity. The ingestion of fermented goat milk also varied

both the preponderance and ratio of lactic acid bacteria species in the gut micro flora of the cases [5].

During metabolic process lactic acid bacteria produce a substance which act against pathogens and that substance is known as bacteriocin. Bacteriocins used in food industry for preserving the food products in natural way [6]. The sources for these probiotic strains are milk, meat and other fermented food products. The aim of the present study was to identify and characterize the novel bacteriocin producing lactic acid bacteria from raw goat milk.

## II. METHODOLOGY

The various raw goat milks were collected from different regions of Tirupur and Erode. The samples were cultured in the De Man Rogosa and Sharpe (MRS) agar media, after the growth of the organisms it were subjected to Gram staining, various biochemical tests. Based on non-haemolytic, antibiotic sensitive pattern, litmus assay and some of the strains were chosen for further probiotic characterization [7]. Such strains were S3, G5, G9, G13, G14, G15, G17, G26, and G27.

**Selection of isolates:** The nine isolates of the lactic acid bacteria identified from various raw goat and sheep milk samples were taken to determine the potential probiotic properties.

#### Cell surface hydrophobicity test [8]

The degrees of hydrophobicity of the strains were determined by employing the method described as in [9]. This method was based on adhesion of cells to hexadecane droplets. Cultures were grown in 10 ml MRS broth, centrifuged at 6,000xg for 5 minutes and the cell pellet was washed and resuspended in 10 ml of Ringer solution 16% NaCl, 0.0075% KCl, 0.01% CaCl<sub>2</sub> and 0.01% NaHCO<sub>3</sub>. The absorbance at 600 nm was measured. Cell suspension was then mixed with equal volume of n-hexadecane and mixed thoroughly by vortexing for 2 minutes. The two phases were allowed to separate for 30 minutes and absorbance at 600 nm of the lower phase was recorded

The percentage hydrophobicity of strain adhering to hexadecane was calculated using the equation.

$$\text{Hydrophobicity (\%)} = \frac{\text{OD}_{600}(\text{initial}) - \text{OD}_{600}(\text{with hexadecane}) \times 100}{\text{OD}_{600}(\text{initial})}$$

#### Phenol resistance tests [10]

Overnight grown active cultures (0.5 ml) were inoculated in MRS broth (5 ml) tube to the concentration of phenol level 0.4 % along with the respective control. The cultures were incubated at 37°C. After 24 hours of incubation extent of growth was recorded by taking the absorbance at 600 nm.

#### Antioxidant Assay

##### DPPH Method [11]

DPPH (Di Phenyl Picryl Hydroxide) activity test in which to a small amount of sample 0.1 ml of DPPH solution was added and mixed well. To that, 400µl of 50mM Tris hydrochloric acid was added and incubated for about 30 minutes at room temperature and the results were determined at 517 nm.

##### Total antioxidant Assay [12]

Take 1 ml of sample to that add 1 ml of the reaction mixture solution which contains the 0.6M H<sub>2</sub>SO<sub>4</sub>, 28 Mm sodium phosphate and 4 Mm ammonium molybdate, mixed well the sample and the reaction mixture were incubated at 50°C for 90 minutes at water bath. After that cool the mixture and the results were determined at 695 nm.

##### Cholesterol reducing activity [13]

From the fresh culture take 1 ml of cell free supernatant and then the volume of the supernatant is made up to 5ml by adding FeCl<sub>3</sub>.CH<sub>3</sub>COOH reagent (0.05%) and 3 ml of concentrated sulphuric acid. After the sample were incubated for 20 minutes at room temperature. Take the absorbance at 560 nm. The control should employed by the medium alone.

#### Aggregation Study

##### Auto aggregation test [14]

The isolates were grown in MRS broth or BHI broth, respectively, for 24 h at 37°C. The cells were harvested by centrifugation (7000 g 10 min 20°C) washed resuspended and diluted in sterile saline water (0.85% NaCl w/v) to obtain an OD<sub>660</sub> nm = 0.3, determined using a spectrophotometer. After incubation for 60 min at 37°C cells were harvested (300 g 2 min 20°C) and the OD<sub>660</sub> nm of the supernatant were assessed accordingly. Auto aggregation was determined using the following equation:

$$\text{Auto aggregation (\%)} = \left[ \frac{\text{OD}_0 - \text{OD}_{60}}{\text{OD}_0} \right] \times 100$$

OD<sub>0</sub> refers to the initial OD, and

OD<sub>60</sub> refers to the OD determined after 60 min.

##### Co-aggregation test [14]

For the evaluation of co-aggregation the isolates were grown in 10 ml of MRS broth at 37°C. Cells were harvested after 24 h (7000 g 10 min 20°C) washed resuspended and diluted in sterile saline water to OD<sub>660</sub> nm = 0.3. The degree of co-aggregation was determined by OD readings obtained for paired studied culture and co-aggregation partners suspensions (ratio of 500 µl and 500 µl of each suspension). Cells were harvested (300 g, 2 min, 20°C), and the OD<sub>660</sub> nm of the supernatant was determined. Coaggregation was calculated using the following equation

$$\text{Co-aggregation (\%)} = \left[ \frac{\text{OD}_{\text{tot}} - \text{OD}_s}{\text{OD}_{\text{tot}}} \right] \times 100$$

OD<sub>tot</sub> refers to the initial OD, taken immediately after the relevant strains were paired.

OD<sub>s</sub> refer to the OD of the supernatant obtained after 60 minutes of incubation.

##### Antimicrobial activity [15]

The agar over lay method was employed to determine the ability of the viable lactic acid bacteria strains to inhibit the growth of the indicator pathogens *E.coli*, *Staphylococcus aureus* and *Candida* spp. A loopful of LAB in MRS broth were inoculated on MRS agar plate as a thick line of about 2 mm and above 30 mm long at a good distance away from the edge of the plates and incubated under microaerophilic condition at 37°C for 24 hours. After incubation, the MRS agar plates were overlaid with approximately 0.2 ml \* 10<sup>7</sup> CFU/ ml of an overnight broth culture of the test pathogens inoculated in 10 ml of the Mueller Hinton Agar (with 0.7% agar-agar). The overlays were allowed to set, and incubated at 37°C under aerobic condition. The plates were then examined for clear zone of inhibition around the line of LAB and the clear zones were measured.

##### Bacteriocin activity [16, 17]

The selected strains were grown in MRS broth at 37°C for 24 hrs. Cell free supernatants were collected by centrifugation (8500 rpm, 10 min, 4°C) of overnight MRS broth cultures. The pH value of the culture supernatant was adjusted to 7

with NaOH (6M) to eliminate the effect of organic acids. After adding catalase (300 U/ml) the cell free supernatants (pH 7) were incubated at 37°C for 1 hr to eliminate the effect of hydrogen peroxide. Then they were heated at 90°C for 10 min to stop the enzyme reaction. The same supernatant without catalase were used as a control and all culture supernatant were filter sterilized to eliminate the possible presence of viable cells (modified).

The inhibition activity was examined by means of the diameters of inhibition zones using the agar well diffusion method. Briefly 10, 15, 20 µl of the cell free supernatant were placed into a wells on the appropriate media agar plates which is seeded with indicator strains *E. coli*, *Streptococcus aureus*. After incubation of 24 hrs the diameters of the inhibitory zones were measured.

#### **Production and Purification of Bioactive compound by using Column chromatography [18]**

The isolates were grown in a production media, MRS media for 24 hrs at 37°C. Then it was centrifuged at 8000 rpm for 10 minutes. 400 g of ammonium sulphate per litre of culture supernatant were added and allowed to settle for 24 hrs at 4°C. The protein precipitate were collected by centrifugation at 6000 rpm/min for 20 minutes and dissolved in 50 ml of 20 mM/L sodium phosphate buffer (pH 6.0). Further it was applied on Diethylamino ethyl-cellulose column (1.5 x 40.0 cm) equilibrated with 0.1 M/L Tris-HCl buffer (pH- 9) and eluted with linear salt gradient of NaCl (0-1 mol/L) the active fractions were pooled together and concentrated by ammonium sulphate loaded on sephadex G-75 column (1.5940 cm) equilibrated with 0.1 mol/L Tris-HCl buffer (pH-9) and eluted with same buffer at a flow rate of 0.5 mL/minute and then eluted highest fractions were subjected to SDS-PAGE.

#### **SDS- PAGE [19]**

An intact SDS PAGE electrophoresis system should include: a tank, lid with power cables, electrode assembly, cell buffer dam, casting stands, casting frames, combs (usually 10-well or 15-well) and glass plates (thickness 0.75mm or 1.0mm or 1.5mm). The SDS PAGE gel in a single electrophoresis run can be divided into stacking gel and separating gel. Stacking gel (acrylamide 5%) is poured on top of the separating gel (after solidification) and a gel comb is inserted in the stacking gel. The acrylamide percentage in SDS PAGE gel depends on the size of the target protein in the sample. The samples were loaded in the gel and after running the samples were subjected to coomassie brilliant blue and the bands were observed.

### **III. RESULTS AND DISCUSSION**

A number of strains of active lactic acid bacteria were isolated from raw goat and sheep milk. The strains which

produce antimicrobial substance were detected by solid culture medium. These strains were screened for potential probiotic characteristics LAB.

#### **Screening of isolates:**

**Cell surface hydrophobicity test** - The degree of hydrophobicity of nine isolates (S3, G5, G9, G13, G14, G15, G17, G26, G27) has been determined and their results were tabulated (Table 1) shown in the figure 1.1. Bacterial surface features is one of the *ex vivo* properties that have been consorted with adherence to a variety of substrate, which in turn it is consorted with hydrophobicity. Bacterial attachment can also be used to influence the colonization potentiality of a microorganism. Through adhesion ability and colonization of tissues, probiotic microorganisms can forbid pathogen approach by steric interactions or particular occlusion on cell receptors [20]. The hydrophobicity of the cell directly proportional to the level of adhesion [21]. Reference [22] was compared and the result that 6 isolates are greater than 50% hydrophobicity which comes under the members of the genus *Lactobacillus*. In our study G5 isolate shows high degree of hydrophobicity and G17 isolate shows low degree of hydrophobicity. In nine isolates 6 strains shows greater than 50% hydrophobicity.

**Phenol resistance test** - These 9 strains showed growth in the presence of 0.4% phenol. Tolerance to phenol is a characteristic feature as phenols can be formed in the intestines by bacteria that de-amine few aromatic amino acids delivered by the diet or produced by endogenous proteins [10]. In our study, the selected 9 strains S3, G5, G9, G13, G14, G15, G17, G26 and G27 were able to tolerate 0.4 % phenol. These 9 strains showed growth in the presence of 0.4%, phenol. G9 shows more tolerance and G26 shows low tolerance. The result were tabulated (Table 1) and shown in the figure 1.2.

**Antioxidant assay**- The DPPH values were measured by the absorbance value at 517 nm for the various nine strains isolated from different raw goat milk samples. The results were tabulated (Table 1) and shown in the figure 1.3a. The DPPH assay is democratic in natural product antioxidant studies, method was mere and sensitive. This assay was founded on the theory that a hydrogen donor is antioxidant. It evaluates the compounds that are radical scavengers [11]. Total antioxidant assay were examine often which is used to approach the antioxidant position of biological sample and can analyze the antioxidant reaction versus the free radicals developed [12]. The total antioxidant assay was measured by the absorbance value at 695 nm for the various nine strains for total antioxidant assay (Figure 1.3b).

**Cholesterol reducing activity** - Several studies have strongly proposed that fermented milk could lower total cholesterol and low density lipoproteins (LDL) cholesterol

thus having a hypocholesterolemic effect [23,24]. The number of cholesterol absorption by these probiotic organisms brought out a broad fluctuation (29 – 57%) quantity of the strains. In another study of *L. yoghurt* has highest (p<0.05) cholesterol absorption capacity while *L. casei* showed moderate and *L. choozti* shows the least [25]. Therefore it is indicated that probiotic can also be used to treat hypercholesterolemic patients, thus reducing the heart attacks. In our study of nine isolates G9 shows high cholesterol reducing activity, G5 shows low cholesterol reducing activity and other isolates shows moderate activity. The cholesterol reducing activity of various isolates were measured at 560nm, the results were tabulated (Table 1) and shown in the figure 1.4.

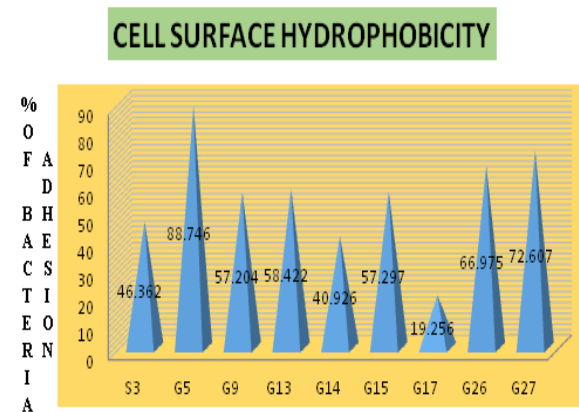
**Table 1. Probiotic Evaluation profile of the isolates**

| S. No | Strain | Antioxidant assay |                   | Phenol resistance test | Cell surface hydrophobicity | Cholesterol reducing activity |
|-------|--------|-------------------|-------------------|------------------------|-----------------------------|-------------------------------|
|       |        | DPPH              | Total antioxidant |                        |                             |                               |
| 1     | S3     | 0.321             | 0.202             | 0.29                   | 46.362                      | 0.278                         |
| 2     | G5     | 0.449             | 0.342             | 0.13                   | 88.746                      | 0.034                         |
| 3     | G9     | 0.270             | 0.418             | 0.28                   | 57.204                      | 0.718                         |
| 4     | G13    | 0.292             | 0.396             | 0.12                   | 58.422                      | 0.416                         |
| 5     | G14    | 0.234             | 0.396             | 0.18                   | 40.926                      | 0.293                         |
| 6     | G15    | 0.437             | 0.189             | 0.13                   | 57.297                      | 0.072                         |
| 7     | G17    | 0.331             | 0.293             | 0.14                   | 19.256                      | 0.276                         |
| 8     | G26    | 0.208             | 0.414             | 0.09                   | 66.975                      | 0.412                         |
| 9     | G27    | 0.444             | 0.542             | 0.25                   | 72.607                      | 0.014                         |

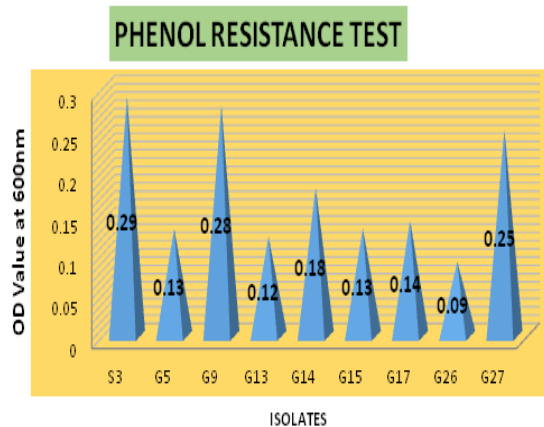
**Auto aggregation and co aggregation-** Based on the probiotic evaluation of G13 was subjected to aggregation study. Sedimentation rate of G13 isolates was determined for a period of 24 hours and the auto aggregation and co aggregation of G13 was measured which shown in the figure 2. Auto aggregation possible of cells acts a significant function in attachment to enteric cells [26] and prevents the infectious agent colonization.

Probiotics should have auto aggregation possibility over 40%. In our study G13 isolates showed affinity over 40% (i.e., 56%). Among 9 isolates, G13 showed highest affinity. This confirms the ability of G13 to adhere, persist and divide in GIT, qualifying it as potential probiotic. Co aggregation

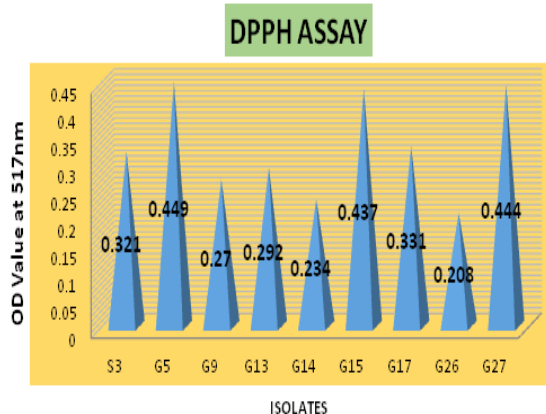
tests plays simple and authentic methods relevant to a large number of the test strains for screening Lactobacillus as described in [27].



**Fig 1.1: Cell surface hydrophobicity test**



**Fig 1.2: Phenol resistance test**



**Fig 1.3a: Antioxidant assay-DPPH**

**TOTAL ANTIOXIDANT ASSAY**

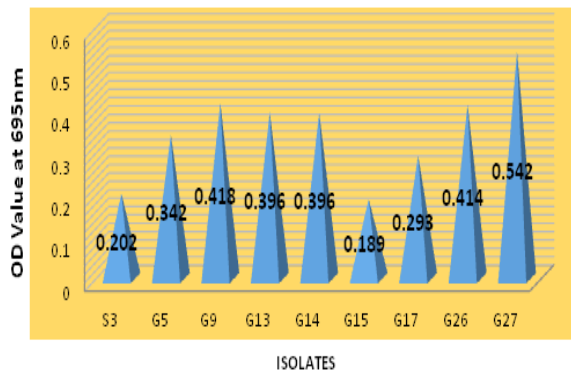


Fig 1.3b: Total antioxidant assay

**CHOLESTROL REDUCING ACTIVITY**

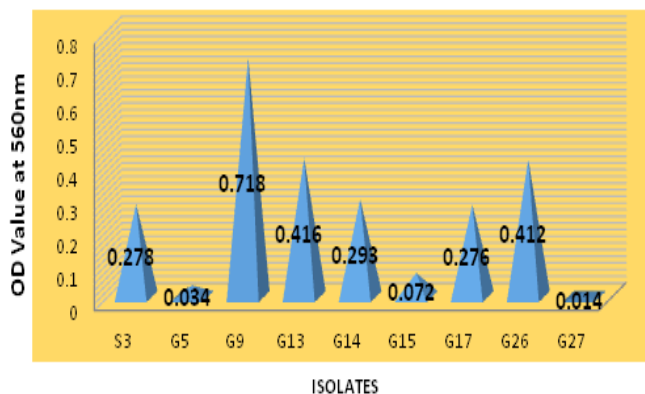


Fig 1.4: Cholesterol reducing activity

Figure 1: Graphical analysis of Probiotic evaluation profile of the goat milk isolates

These attributes are believed to be associated to the ability to act intimately with unsuitable bacteria. Thus in study of the probiotic from chicken gut, the organism *Lactobacillus salivarius* gives high coaggregation [28]. In our study the isolate G13 gives 35.75% when interact with *E.coli* and 57.05% when interact with *Staphylococcus aureus*.

**Antimicrobial activity**-The diameters of inhibition zones showed that all the isolates have antibacterial effects against the tested pathogens were tabulated (Table 2) and shown in the figure 3. Antimicrobial activity against pathogens desolating goat production is a suitable attributes of a probiotic strain to be used in goat's farming. Antibacterial activity is life sustaining for the successful colonization of lactobacilli in the intestinal mucosa as they allow a barrier effect and defence against pathogens [29]. *Lactobacillus* may obtain antimicrobial effect by producing some substances such as organic acids, ( lactic, acetic, propionic) carbon dioxide, hydrogen peroxide, diacetyl, low molecular weight

antimicrobial substances such as bacteriocins, which may be endlessly excreted by the bacteria to produce the inhibitor activity versus the pathogens [30].

**ANOVA Analysis for Aggregation test**

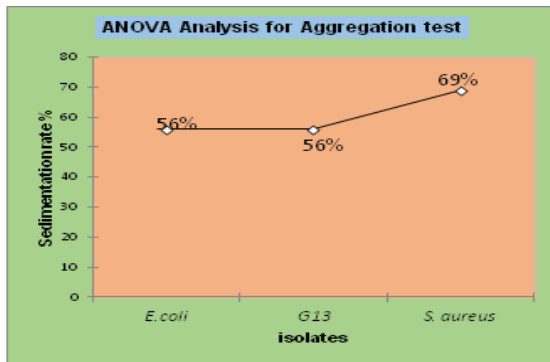


Fig 2.1: Auto aggregation assay

**ANOVA Analysis for Co-aggregation Assay**

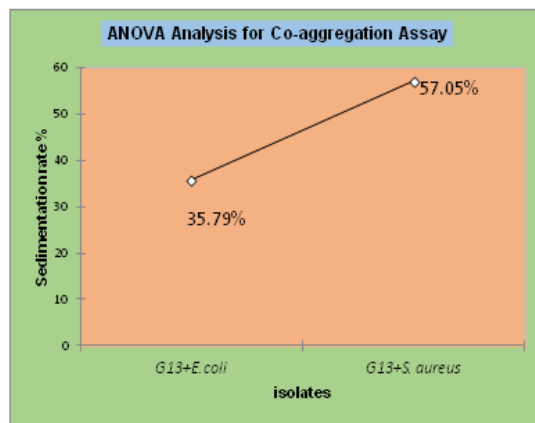


Fig 2.2: Co-aggregation assay

Figure 2: ANOVA analysis for Aggregation Study

The antibacterial activity of the cell free supernatants prevailed from the G5 and G13 probiotic isolates were tested by agar well diffusion method against *E. coli*, *S. aureus* and *Candida albicans* as selected pathogens of importance in goat's farming. The antimicrobial activity produced by the 2 isolates in this study demonstrated inhibition of growth for *E. coli*, *S. aureus* and *C. albicans*. The two isolates have inhibition zones with diameter of 7mm and 9mm against *S. aureus* for G5 and G13, no zone and 7mm against *E. coli* and show no zone for G5 and G13 against *C. albicans*.

Table 2: Antimicrobial activity

| Organisms                    | G5      | G13     |
|------------------------------|---------|---------|
| <i>E.coli</i>                | 10 mm   | 14 mm   |
| <i>Staphylococcus aureus</i> | 9 mm    | 8 mm    |
| <i>Candida albicans</i>      | No Zone | No zone |



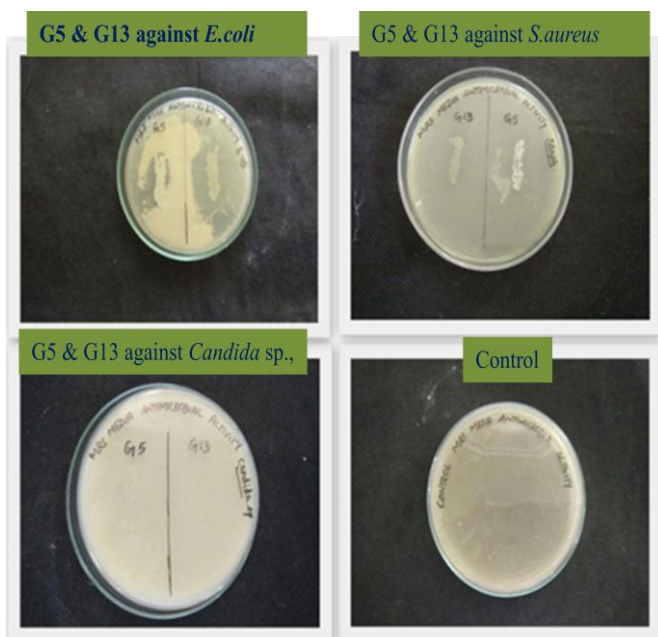


Figure 3:Plates of Antimicrobial activity

**Column chromatography** -Using column chromatography were the protein purified from the G13 isolate, their fractions were tabulated (Table 4.11.) and the highest fraction has taken for SDS-PAGE analysis to know the molecular weight of the protein produced by G13 isolate in the crude.

Table 3: Fraction obtained from chromatography

| S.NO | FRACTIONS (280nm) |
|------|-------------------|
| 1    | 0.239             |
| 2    | 0.430             |
| 3    | <b>0.452</b>      |
| 4    | 0.381             |
| 5    | 0.436             |

**Cell-free supernatant antibacterial assay**-The antibacterial activity of the cell free supernatants obtained from the isolates were tested by agar well diffusion method against *E. coli* and *S. aureus*. In this study manifested inhibition of growth for *E. coli* and *S. aureus*. The inhibitory agents produced by the isolated LAB examined in this study could be characterized as bacteriocins like, since inhibition due to acid have been excluded. Results shown in table 4. The well diffusion method was done to determine the bacteriocin activity of the LAB cultures. The bacteriocin produced and was purified and the sensitivity to test organism were carried out thus in the study *L.plantarum* and *L.brevis* shows high inhibition against the *E.coli* and *Salmonella* sp [31].

Table 3.Cell free crude antibacterial agar well diffusion assay

| Test Organism                | 10 μL | 15 μL | 20 μL | pH7 10μ L | pH7 15μ L | pH7 20μ L | MRS contrl | MRS pH7 contro l |
|------------------------------|-------|-------|-------|-----------|-----------|-----------|------------|------------------|
| <i>E.coli</i>                | 4 m m | 6 m m | 6 m m | 4 mm      | 5 mm      | 6 mm      | No zone    | No zone          |
| <i>Staphylococcus aureus</i> | 5 m m | 6 m m | 7 m m | No Zone   | 4 mm      | 7 mm      | No zone    | No zone          |

**SDS -PAGE** - Considering bacteriocin production, cell growth was started from late log phase itself and maximum was obtained in early stationary growth phase at 30th hr of the culture. Growth beyond the stationary phase resulted decrease in bacteriocin production. In purification bacteriocin from culture supernatant was concentrated by ammonium sulphate precipitation followed by cation exchange chromatography and hydrophobic interaction chromatography. From this study the five fraction were collected and the highest fraction 0.452 were screened. Single protein bands were observed when stained with coomassie brilliant blue and it clearly indicated the purity of the protein. SDS-PAGE analysis showed an electrophoretically pure protein with an apparent molecular size of 17 kDa which shows the present of low molecular weight protein. The molecular weight of the purified bacteriocin of G13 isolates was estimated at 17 kDa (Figure 4) by SDS- PAGE.

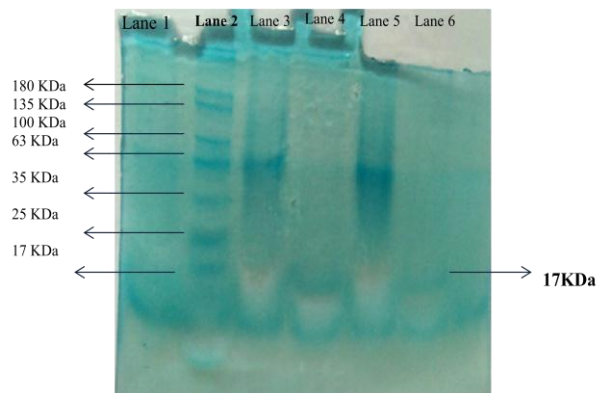
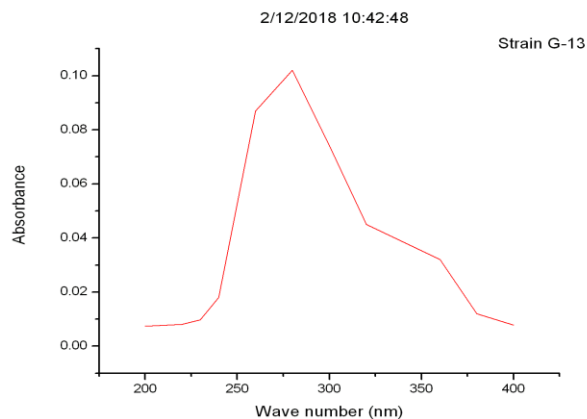


Figure 4: Molecular weight of the purified bacteriocin from G13 by SDS-PAGE.

# Note: lane 1 - blank, lane 2-marker, lane 3-crud of other isolate, lane 4-band of other isolate purified sample, lane 5-crude of G13 and lane 6- G13 isolates purified sample.

**UV-Visible spectrometric analysis-** Bacteriocins produced by Lactic acid bacteria were come under 10KDa such as nisin (3,5 KDa), lactisin(4,5KDa), lactacin(6,8KDa), Lactocin (3,7 KDa), sakacin P (4,4KDa) and Helveticin (<30 KDa) reported in [32] and reported *L.murinus* AU06 produced 21KDa protein[18]. In our study purified protein from G13 antibacterial compound molecular weight 17 KDa. It may be a novel protein produced by lactic bacteria.

According to the UV-VIS Spectrophotometry analysis (Figure 5) our sample showed 280 nm, so they were proteinaceous nature compound. Protein absorb in 240 - 300nm based on the presence of amino acid [33].



**Figure 5: Microprocessor UV-VIS SPEC single beam analysis of band of G13 sample from SDS-PAGE**

#### IV. CONCLUSION AND FUTURE SCOPE

The present study has concluded that most of the strains isolated from raw goat and sheep milk possess good probiotic characteristic. G13 isolate producing antimicrobial compound showed 17KDa molecular weight protein. It may be a novel protein, needs further compound analysis. Further work required to carry out safety zone measurement of using probiotic bacteria in food industry.

#### Conflict of interest

None

#### ACKNOWLEDGMENT

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